## Is the similarity significant, or could it be due to chance?

Even if two proteins are unrelated, we would expect some similarity simply by chance.

Is the alignment score significantly higher than random?

Align random permutations of the sequences, and find the mean and standard deviation of the resulting distribution.

The z-score reflects the significance of a global similarity score.

z-score =  $\frac{score - mean}{score}$ standard deviation

Larger values imply greater significance.

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## More realistic similarity measures

Not all substitutions are equally likely.

- A transition between two purines (A, G) or between two pyrimidines (C, T/U) is more common than a purine-pyrimidine transversion.
- Replacement of one amino acid residue by another with similar size or physiochemical properties is more common than replacement by a dissimilar amino acid residue.

Insertion/deletion of N contiguous amino acid residues or nucleotides is more likely than N independent insertion/deletion events.

Thus, we should have different penalties for opening gap and for extending a gap.

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## Possible substitution matrices for DNA

	Α	С	G	Т		
Α	2	-1	-1	-1		
С	-1	2	-1	-1		
G	-1	-1	2	-1		
т	-1	-1	-1	2		
•		•	•	-		
	Δ	C	G	т		
Δ	2	-2	_1	-2		
	2	-2	-1	-2		
Č	-2	2	-2	-1		
G	-1	-2	2	-2		
I	-2	-1	-2	2		
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## Substitution matrices

PAM and BLOSUM are two widely used families of substitution matrices.

Derived from observations of actual mutations in alignments of related proteins.

observed ab mutation rate

 $score_{ab} = \log \frac{1}{mutation rate expected from amino acid frequencies}$ 

PAM and BLOSUM differ in how the alignments, from which observations are made, are selected.

Substitution matrices near one end of the series are most suitable when comparing proteins separated by a short evolutionary distance; those near the other end are most suitable for comparing distantly related proteins.



## BLOSUM62 A R N D C O E G H I L K M F P S T W Y V A 4 R -1 5 N -2 0 6 D -2 -2 1 6 C 0 -3 -3 -3 9 0 -1 1 0 0 -3 5 E -1 0 0 2 -4 2 5 G 0 -2 0 -1 -3 -2 -2 6 H -2 0 1 -1 -3 0 0 -2 8 I -1 -3 -3 -3 -1 -3 -3 -4 -3 4 L -1 -2 -3 -4 -1 -2 -3 -4 -3 2 4 к -1 2 0 -1 -3 1 1 -2 -1 -3 -2 5 M -1 -1 -2 -3 -1 0 -2 -3 -2 1 2 -1 5 F -2 -3 -3 -3 -2 -3 -3 -3 -1 0 0 -3 0 6 P -1 -2 -2 -1 -3 -1 -1 -2 -2 -3 -3 -1 -2 -4 7 S 1 -1 1 0 -1 0 0 0 -1 -2 -2 0 -1 -2 -1 4 т 0 -1 0 -1 -1 -1 -1 -2 -2 -1 -1 -1 -1 -2 -1 1 5 W -3 -3 -4 -4 -2 -2 -3 -2 -2 -3 -2 -3 -1 1 -4 -3 -2 11 Y -2 -2 -2 -3 -2 -1 -2 -3 2 -1 -1 -2 -1 3 -3 -2 -2 2 7 V 0 -3 -3 -3 -1 -2 -2 -3 -3 3 1 -2 1 -1 -2 -2 0 -3 -1 4 Graham Kemp, Chalmers University of Technology

# Structural clues from multiple sequence alignments — Residues at highly conserved positions often have important functional or structural roles. — Insertions and deletions can be accommodated most easily in surface loops. — Conserved patterns of hydrophobic residues can suggest secondary structure. — The root mean square deviation between pairs of homologous proteins generally increases as the percent residue identity decreases. Gratum Name, Chatmens University of Technology

- Many α-helices are amphipathic. Conserved hydrophobic residues at positions i, i+3, i+4, i+7, etc. are highly indicative of an α-helix.
- Half-buried strands will tend to have hydrophobic and hydrophilic residues at alternate positions.
- In proteins containing both α-helices and strands the strands are often completely buried and tend to contain only hydrophobic residues.

For more details and references, see:

http://www.bmm.icnet.uk/people/rob/CCP11BBS/secstrucpred.html





## "HSSP-curve"

- Shows the length-dependent threshold for significant sequence identity.
- Proposed by Sander and Schneider (1991) and revised by Rost (1999).
- Above the curve, identifing true positives is easy.
- Just below the curve, the number of false positives rises rapidly; distinguishing between true and false positives in the "twilight zone" is difficult.

(HSSP stands for "Homology-derived Secondary Structure of Proteins")



## Comparative modelling and fold recognition

Comparative modelling (homology modelling): Given:

- sequence of target protein with unknown structure
- known structure of a related protein

Predict:

• three-dimensional structure of target protein

Fold recognition:

Given:

- sequence of target protein with unknown structure
- library of known folds

Predict:

known fold that is most compatible with the target protein's sequence

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# Comparative modelling strategy

- identify a known structure that is predicted to be similar;
- align sequences;
- predict structurally conserved regions, and locations of insertions and deletions (sometimes called "indels");
- build model backbone structure
  - copy predicted conserved main chain regions from template structure,
  - remodel loops with insertions or deletions;
- add side chains to the modelled main chain;
- evaluate and refine model.

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## **Fold recognition**

The idea behind "threading":

Imagine a wire wound into the shape of a known protein's main chain "fold".

Imagine next that our new sequence is represented by beads that are "threaded", in order, onto the wire, and are pushed along the wire.

At each step, a score is calculated based on which residues are adjacent in space, which residues are buried, etc.

Repeat this process for each different known fold.

A high score indicates that the sequence is compatible with that fold.

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# Further studies in bioinformatics

Sequence MVE360 - Bioinformatics

Structure TDA507 - Computational methods in bioinformatics

Systems KMG060 - Systems biology