

### Why build model structures?

Knowledge of a protein's three-dimensional structure is vital to a full understanding of the molecular basis for its biological function.

We want to understand the function of all proteins encoded by a genome, therefore we would like to know all of their 3-D structures.

Experimental techniques for determining protein structure are relatively slow and expensive, so we look to modelling as a way of extending the set of 3-D structures.

Modelling can also be used in protein engineering when designing proteins for therapeutic applications.

### Using known substructures in protein crystallography

Jones, T.A. and Thirup, S. (1986)  
The EMBO Journal, vol. 5, pp 819-822.

Electron density map interpretation is made easier by fitting regular  $\alpha$ -helices and strands into the map.

This building-block approach to protein modelling can be extended to include **all** main chain fragments.

For example, a model of retinol binding protein was built using fragments from only three other proteins. A model with  $C\alpha$  atoms matching within an R.M.S. error of 1Å was built using only 15 fragments.

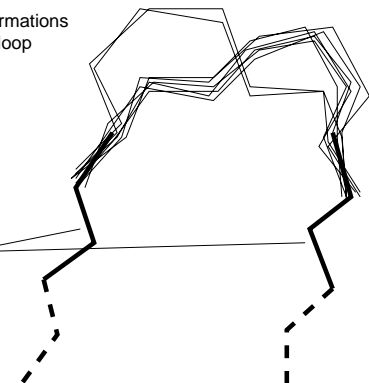
### Comparative modelling strategy

- identify a known structure that is predicted to be similar;
- align sequences;
- predict structurally conserved regions, and locations of insertions and deletions (sometimes called "indels");
- build model backbone structure
  - copy predicted conserved main chain regions from template structure,
  - remodel loops with insertions or deletions;
- add side chains to the modelled main chain;
- evaluate and refine model.

### Fragment-fitting: an approach to remodelling loops

Candidate conformations  
for replacement loop

Structurally  
conserved  
framework



### Fragment selection criteria

- steric overlap;
- packing
  - no protruding loops;
  - no internal cavities;
- disulphide bridges and salt bridges;
- solvent accessibility
  - avoid burying unpaired charges;
- sequence criteria
  - Gly and Pro residues
  - similarity between model's sequence and the sequences of the fragments in their native structures.

### Side chain rotamers

There is an extremely large number of possible combinations of side chain conformations — infinite if we consider side-chain bonds to be continuously variable.

For practical purposes the search space can be discretised by considering a finite set of possible torsion angles for each side-chain.

The distribution of side chain conformations falls into statistically significant clusters. By using representative side chain conformations, or **rotamers**, the vast combinatorial search space can be greatly reduced.

Ponder, J.W. and Richards, F.M. (1987)  
J. Mol. Biol., vol. 193, pp 775-791.

### Cluster fragments

Kelley, L.A., Gardner, S.P. and Sutcliffe, M.J. (1996)  
An Automated Approach For Clustering An Ensemble Of NMR-Derived Protein Structures Into Conformationally-Related Subfamilies. Protein Engineering, vol. 9, pp 1063-1065.

RMS distances between pairs of structures or fragments.

Clusters built using average linkage.

Penalty function seeks to simultaneously minimise:

- a) the number of clusters;
- b) the spread across each cluster.

### Energy calculations

Terms used in evaluating the energy of a conformation typically include:

- bond stretching
- bond angle bend
- terms penalising deviation from planarity, etc.
- torsion angles
- Van der Waals interactions
- hydrogen bonds
- electrostatics
- interactions with solvent, water and cosolutes